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GERMACRANOLIDES FROM *ERLANGEA CORDIFOLIA**

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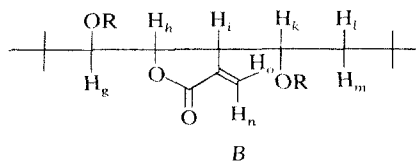
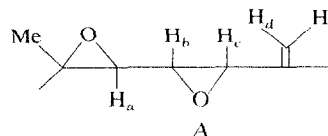
Key Word Index—*Erlangea cordifolia*; Compositae; Vernonieae; new sesquiterpene lactones; germacranolides; triepoxide.

INTRODUCTION

So far only 3 members of the large African genus *Erlangea* (Compositae, tribe Vernonieae) have been investigated. In addition to simple sesquiterpenes, one species contains two guaianolides [1], while in a second one germacranolide of the glaucolide type was found [2]. The third species only contains special 5-methylcoumarins [3] also present in *Bothriocline* [4] and *Ethulia* [5], both being African genera belonging to the Vernonieae. A germacranolide is reported in a patent application from *E. cordifolia* [6], however, no stereochemistry was given. We now report the isolation and structure elucidation of two highly oxygenated germacranolides from the same plant, which are named cordifene and 4,15-epoxy-4,15-dihydro-cordifene.

RESULTS AND DISCUSSION

The polar fractions of the aerial parts of *E. cordifolia* afforded, as the main constituent, a crystalline compound with the molecular formula $C_{20}H_{24}O_7$. The 1H NMR spectrum (Table 1) was very similar to the spectrum reported by Mugo [6]. The 270 MHz 1H NMR spectrum displayed typical signals for a methylene lactone (*s(br)*, 6.31 and 5.67) and those for an angelic acid ester (*qq*, 6.08; *dq*, 1.93 and *dq*, 1.79). Extensive double resonance experiments lead to the sequences A and B.



Irradiation of the signal for H_b (*dd*, 3.00) collapsed the doublet at 2.60 and the triplet doublet at 3.97 to singlets. Irradiation at 3.97 sharpened the signals of the methylene protons at 5.53 and 5.39. The coupling J_{H_b, H_c} was 7.5 Hz indicating a *cis* configuration of 1- and 2-H as in mikanolide [7, 8]. As the signal at 3.60 showed no further couplings and as there was a downfield methyl singlet at 1.58, the position of this group seems to be established. On irradiation of H_h (*d(br)*, 4.38), a doublet at 4.12 H_g changes to a singlet, while irradiation of H_i sharpened the broadened singlets at 6.31 and 5.67 (13-H), and the *ddd* 5.11 became a *dd*. This clearly showed that the signal at 2.92 must be assigned to 7-H. Further irradiation of H_k (*ddd*, 5.11) collapsed the two double-doublets at 2.57 and 1.34 to doublets. These results are only in agreement with part B. However, the relative positions of the oxygen functions remained to be elucidated. On acetylation a monoacetate was obtained. In the 1H NMR spectrum (Table 1) the signal

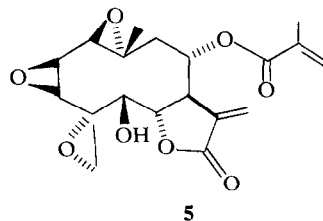
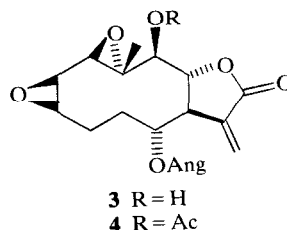
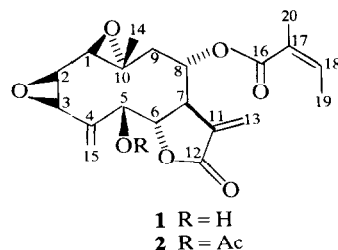
*Part 267 in the series: "Naturally Occurring Terpene Derivatives". For Part 266 see: Balbaa, S. I., Halim, A. F., Halaweish, F. T. and Bohlmann, F. *Phytochemistry* **19**, (in press).

Table 1. ^1H NMR data of compounds **1**, **2** and **5** (270 MHz, TMS as internal standard)

| | 1 | Δ^* | 2 | 5 |
|---------------|-------------------|------------|-------------------|-------------------|
| 1-H | 2.60 <i>d</i> | 0.44 | 2.60 <i>d</i> | 2.63 <i>d</i> |
| 2-H | 3.00 <i>dd</i> | 0.37 | 3.05 <i>dd</i> | 3.06 <i>dd</i> |
| 3-H | 3.97 <i>dt</i> | 0.29 | 3.80 <i>d(br)</i> | 3.83 <i>d(br)</i> |
| 5 α -H | 4.12 <i>d</i> | 0.25 | 5.39 <i>d</i> | 3.23 <i>d(br)</i> |
| 6 β -H | 4.38 <i>d</i> | 0.31 | 4.46 <i>d</i> | 4.53 <i>d</i> |
| 7 α -H | 2.92 <i>d(br)</i> | 0.21 | 2.95 <i>d(br)</i> | 2.93 <i>d(br)</i> |
| 8 β -H | 5.11 <i>ddd</i> | 0.22 | 5.10 <i>ddd</i> | 5.14 <i>ddd</i> |
| 9 α -H | 1.34 <i>dd</i> | 0.24 | 1.34 <i>dd</i> | 1.43 <i>dd</i> |
| 9 β -H | 2.57 <i>dd</i> | 0.18 | 2.59 <i>dd</i> | 2.63 <i>dd</i> |
| 13-H | 5.67 <i>s(br)</i> | 0.10 | 5.70 <i>s(br)</i> | 5.67 <i>s(br)</i> |
| 13'-H | 6.31 <i>s(br)</i> | 0.14 | 6.37 <i>s(br)</i> | 6.30 <i>s(br)</i> |
| 15-H | 5.29 <i>s(br)</i> | 0.12 | 5.44 <i>s(br)</i> | 3.29 <i>d</i> |
| 15'-H | 5.53 <i>s(br)</i> | 0.24 | 5.63 <i>s(br)</i> | 2.73 <i>d(br)</i> |
| 14-H | 1.58 <i>s</i> | 0.16 | 1.60 <i>s</i> | 1.64 <i>s</i> |
| 18-H | 6.08 <i>qq</i> | 0.04 | 6.09 <i>qq</i> | 6.11 <i>qq</i> |
| 19-H | 1.93 <i>dq</i> | 0.04 | 1.93 <i>dq</i> | 1.94 <i>dq</i> |
| 20-H | 1.79 <i>dq</i> | 0.04 | 1.78 <i>dq</i> | 1.80 <i>dq</i> |
| —OH | 3.15 <i>s(br)</i> | 0.22 | — | 3.14 <i>s(br)</i> |
| OAc | — | — | 2.16 <i>s</i> | — |

* Δ values after addition of $\text{Eu}(\text{fod})_3$.

J (Hz): 1,2 = 7.5; 2,3 = 4.5; 3,15 ~ 1; 5,6 = 9.5; 7,8 = 10; 7,13 ~ 1; 8,9 α = 12; 8,9 β = 4; 9 α ,9 β = 12.5; 18,19 = 7.5; 18,20 = 19,20 = 1.5; bei **5**: 15,15' = 5; 15',6 ~ 1.



for H_8 was shifted downfield. This indicated that we were dealing with a germacranolide. However, the stereochemistry and the relative position of the oxygen function still was not clear. It was obvious from the NMR data that the couplings $J_{6,7}$ and $J_{7,8}$ must be very small. Inspection of Dreiding-models showed that we were most probably dealing with a 6,12-*trans*-fused lactone in a conformation with C-14 and C-15 above the plane. The other observed couplings would be in agreement with **1**. However, the second possibility would be **3**. Considering the position of the signal for proton at the OH-bearing carbon, an allylic position as in **1** seemed to be more probable. Furthermore, the observed shifts on acetylation indicated this position as there was a pronounced shift of the signals of the methylene protons (15-H). All efforts to transform the alcohol to the corresponding ketone were unsuccessful. MnO_2 as well as pyridine chlorochromate gave no identifiable products. The stereochemistry at C-5, C-6 and C-8 followed from the observed couplings, while that at C-10 remained uncertain. We have assumed this configuration by analogy to similar lactones. All data therefore were in good agreement with the proposed structure **1**. Also the ^{13}C NMR data (Table 2) supported this assumption. The assignments of most of the signals were easy. However, some may be interchangeable. The unusual positions of the signal of C-8 is probably due to the shielding effect of the C-10 methyl, which support the configuration at C-10.

A second slightly more polar lactone, molecular formula $\text{C}_{20}\text{H}_{24}\text{O}_8$, showed a very similar ^1H NMR spectrum to **1**. The main difference was the absence of the vinylic proton signals for 15-H. They were replaced by a doublet at 3.29 and a broadened doublet at 2.73.

The last signal showed a W-coupling with 5-H. The positions of these signals and the coupling constant indicated the presence of an epoxide of **1**. Though the configuration of this new group could not be established, the NMR data were in good agreement with the α -epoxide **5**. The observed shift of the signal for 6-H if compared with that in **1** again showed that the general structure **1** should be favoured, since it could not be explained by an epoxide of **3**. Therefore, most probably the second lactone was 4,15-epoxy-4,15-dihydrocoidifene (**5**).

The isolation of **1** and **5** from an *Erlangea* species again shows that highly oxygenated sesquiterpene lactones are characteristic for this genus. However, these compounds are also present in the large genus *Veronica*. The chemotaxonomic situation in this tribe is

Table 2. ^{13}C NMR chemical shifts of compound **1** (CDCl_3)

| | | | |
|------|-----------------|------|----------------|
| C-1 | 52.4 <i>d</i> * | C-11 | 134.0 <i>s</i> |
| C-2 | 54.2 <i>d</i> * | C-12 | 167.8 <i>s</i> |
| C-3 | 62.4 <i>d</i> | C-13 | 127.1 <i>t</i> |
| C-4 | 139.5 <i>s</i> | C-14 | 118.6 <i>t</i> |
| C-5 | 80.6 <i>d</i> | C-15 | 17.8 <i>q</i> |
| C-6 | 77.8 <i>d</i> | C-16 | 166.0 <i>s</i> |
| C-7 | 47.9 <i>d</i> | C-17 | 127.0 <i>s</i> |
| C-8 | 67.9 <i>d</i> | C-18 | 140.0 <i>d</i> |
| C-9 | 45.4 <i>t</i> | C-19 | 15.8 <i>q</i> |
| C-10 | 55.2 <i>s</i> | C-20 | 20.1 <i>q</i> |

*Possibly interchangeable.

still not very clear, but as pointed out by Jones [9] the systematic situation is also confused. Perhaps further chemical investigations may be helpful.

EXPERIMENTAL

IR: CHCl_3 ; ^1H NMR: 270 MHz, TMS as int. standard; MS: 70 eV; optical rotation: CHCl_3 . The air-dried plant material (29 g aerial parts) was extracted with Et_2O at room temp. Saturated hydrocarbons were removed by treating with MeOH. The resulting extract was first separated by CC (Si gel, act. grade II). With Et_2O **1** and **5** were eluted, which could be separated by repeated TLC (Si gel GF 254) (Et_2O -petrol, 7:3) yielding ca 30 mg **1** and 15 mg **5**.

Cordifene (1). Colourless crystals from Et_2O -petrol, mp 183° . IR $\lambda_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3580 (OH), 1780 (γ -lactone), 1720 ($\text{C}=\text{CCO}_2\text{R}$); MS m/e (rel. int.): 376 (M^+ 0.6%) $\text{C}_{20}\text{H}_{24}\text{O}_4$; 361 ($-\text{Me}$, 0.3); 358 ($-\text{H}_2\text{O}$, 1); 276 ($-\text{C}_4\text{H}_7\text{CO}_2\text{H}$, 2); 83 ($\text{C}_4\text{H}_7\text{CO}^+$, 100); 55 (83-CO, 70).

$$[\alpha]_{24}^{\text{D}} = \frac{589}{-127.1} - \frac{578}{-133.9} - \frac{546}{-154.8} - \frac{436}{-274.2} \quad (c = 0.31).$$

5 mg **1** in 0.5 ml Ac_2O was heated for 30 min at 70° . After evapn the residue was purified by TLC (Et_2O -petrol, 7:3) yielding 3 mg of **2**, colourless oil, IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1800 (lactone), 1760, 1230 (OAc), 1730 ($\text{C}=\text{CCO}_2\text{R}$), MS (Cl, isobutane m/e (rel. int.): $\text{M}+1$ —; 391 ($-\text{CO}$, 100); 377 ($-\text{ketene}$, 49); 289 ($-\text{CHO}$, $\text{C}_4\text{H}_7\text{CO}_2\text{H}$, 30); 229 (289-HOAc, 17). Oxidation of **1** with MnO_2 or pyridine chloro-

chromate led to degradation of the lactone. Definite products could not be isolated.

4,15-Epoxy-4,15-dihydrocordifene (5). Colourless gum, IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3580 (OH), 1780 (lactone), 1720 ($\text{C}=\text{CCO}_2\text{R}$); ^1H NMR see Table 1.

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